

# Design Principles of a Bacterial Signalling Network

Why chemotaxis is more complicated than needed

Jens Timmer

Center for Systems Biology  
Center for Data Analysis and Modeling  
Center for Applied Biosciences  
Bernstein Center for Computational Neuroscience  
Department of Mathematics and Physics  
University of Freiburg

<http://www.fdm.uni-freiburg.de/~jeti/>

# Outline

- **Introduction**
- **Chemotaxis**
- **Barkai/Leibler Model**
- **Fluctuations, Cell-to-Cell Variability**
- **Design Principles of Robustness**

# Enlarging Physics, Math, Engineering

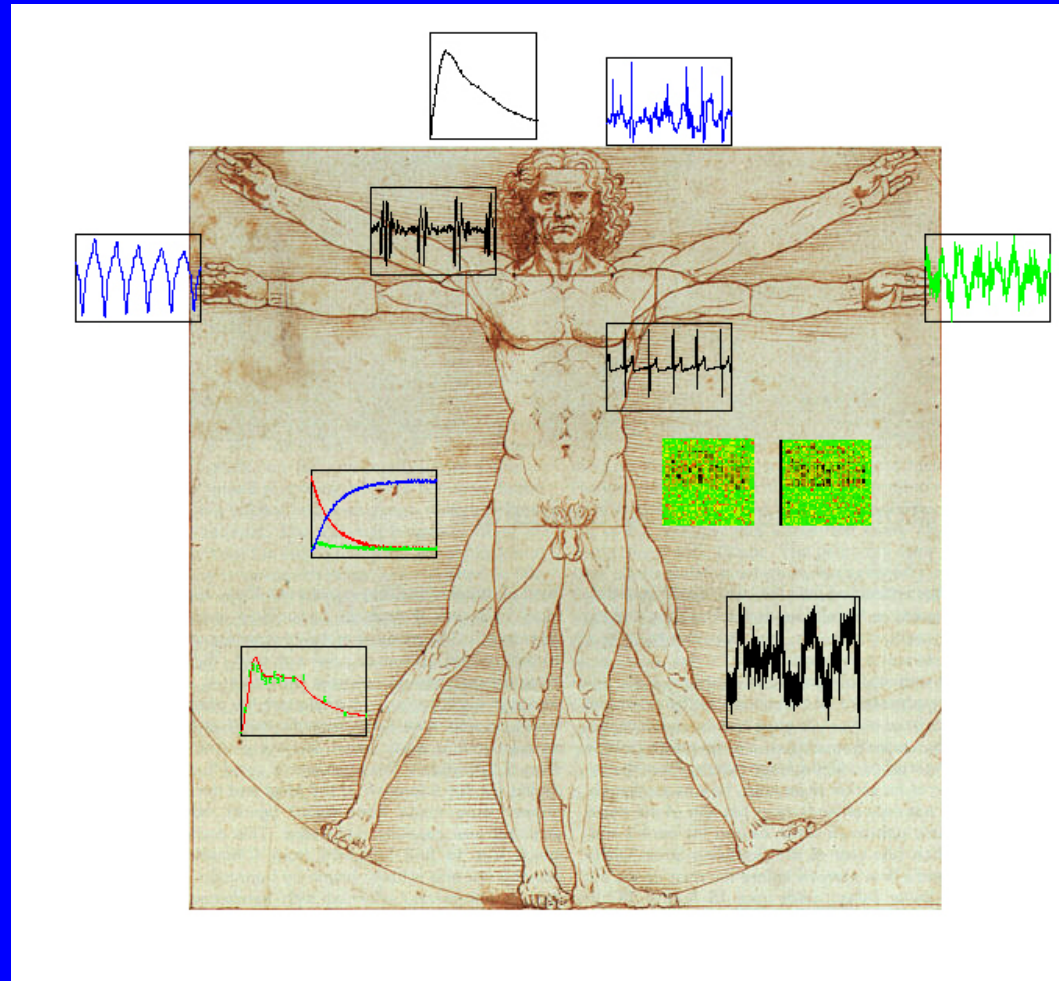
- **Since Newton:**

**Mathematization of inanimate nature**

- **21st century:**

**Additionally: Mathematization of animate nature**

# Man : A Dynamical System



**Diseases caused or expressed by malfunction of dynamical processes**

# Systems Biology

**Understanding biomedical systems by data-based mathematical modelling of their dynamical behavior**

**Based on but more than ...**

- ... **Mathematical Biology: Data-based**
- ... **Bioinformatics: Dynamics**
- ... **o.p./g. – o.p.: System**
- ... **another omics: Mathematics**

# Why Modelling in Cell Biology?

- **Basic Research**

- Genomes are sequenced, but ...
- ... function determined by regulation
- Regulation = Interaction & Dynamics
- Function: Property of dynamic network
- "Systems Biology"

- **Application**

- Drug development takes 10 years and 1 bn \$/€
- Reduce effort by understanding systems

## Two Differences between Physics and Biology

- Fundamental laws of nature vs. principles
- In biology there is "function" due to evolution

Physics in biology:

Apply mathematics to understand function

# Bacterial Chemotaxis – The Phenomenon

- Bacteria sense nutrient gradients over four orders of magnitude of absolute concentration
- Detect relative changes of 2 %
- Robust against perturbations

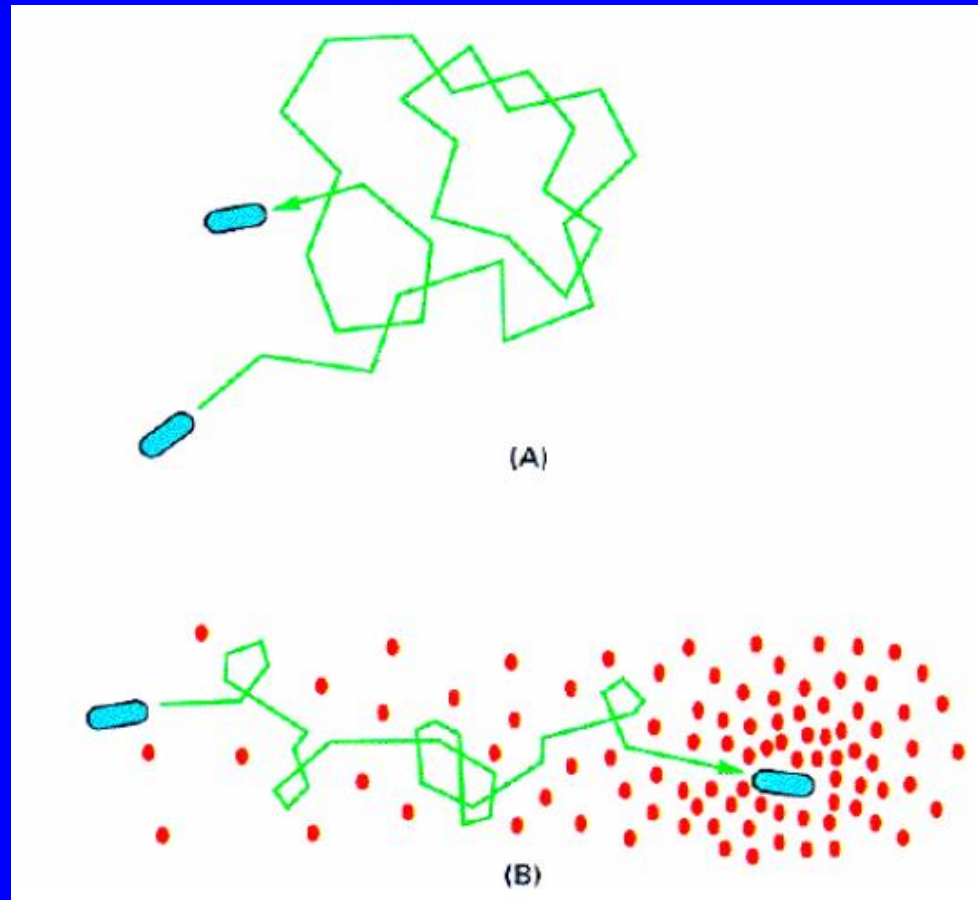
Chemotaxis: One of the best investigated biological systems



# Bacterial Chemotaxis – The Strategy

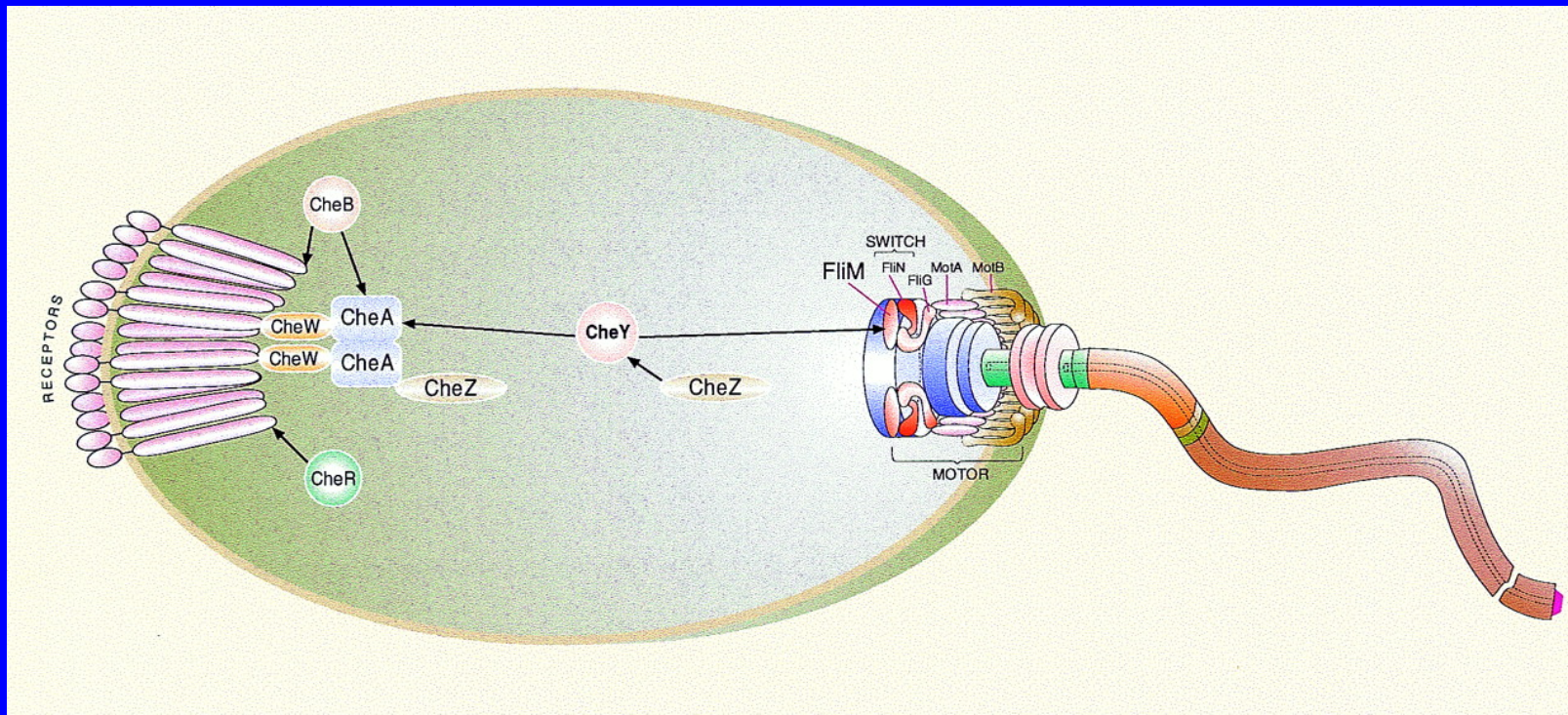
- Bacteria too small to compare front to end
- Strategy:
  - Change direction from time to time (tumble)
  - If concentration increases: reduce tumbling frequency
  - If concentration decrease: increase tumbling frequency
- Sense spatial gradients by temporal changes

# Chemotaxis – Tumble and Swim



Random walk vs. biased random walk

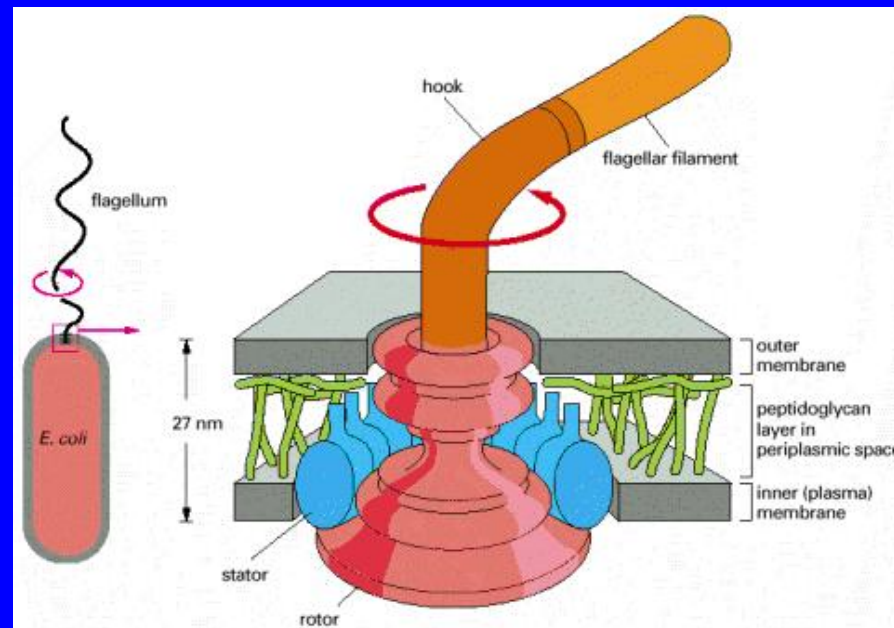
# Chemotaxis in *E. coli*



# Chemotaxis – Flagella

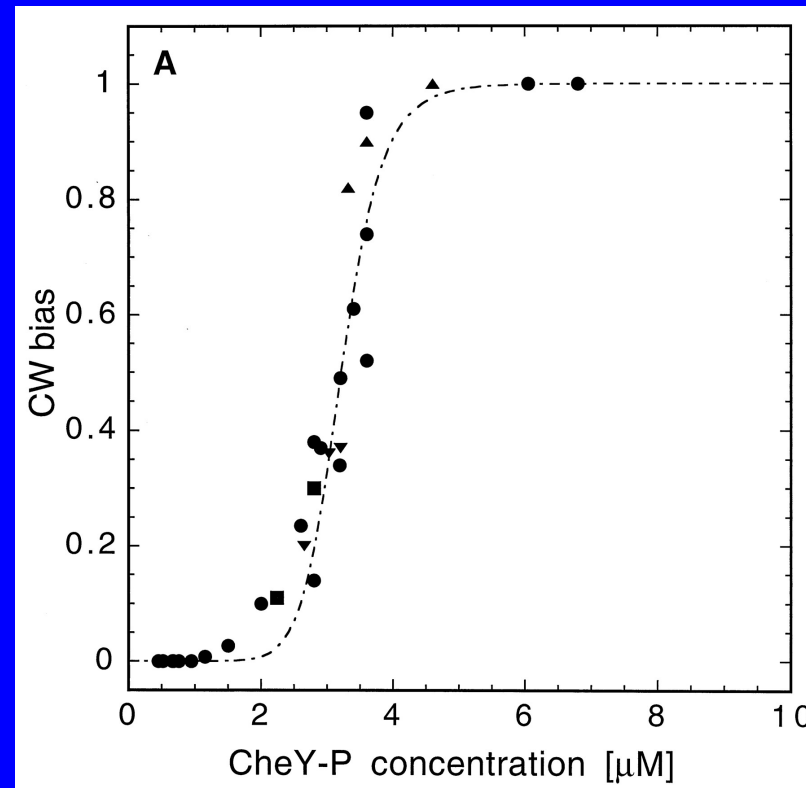
## Movement by rotating corkscrew-flagella

- counter-clockwise: form bundle: swim by marine propeller
- clockwise: rotate radially: tumble



# Chemotaxis – The Task

Tumbling/Swimming depends on phosphorylated CheY



**Important: A small working range**

## Chemotaxis – Adaptation

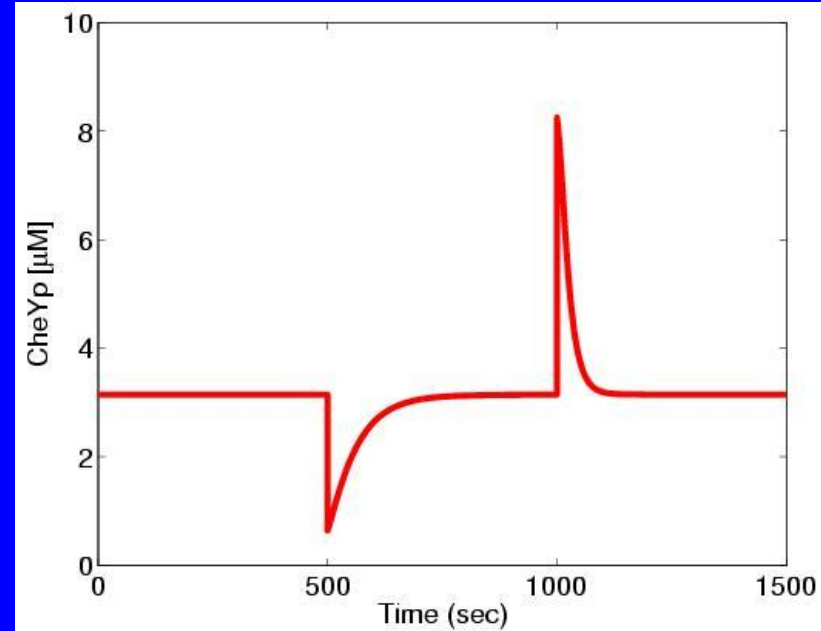
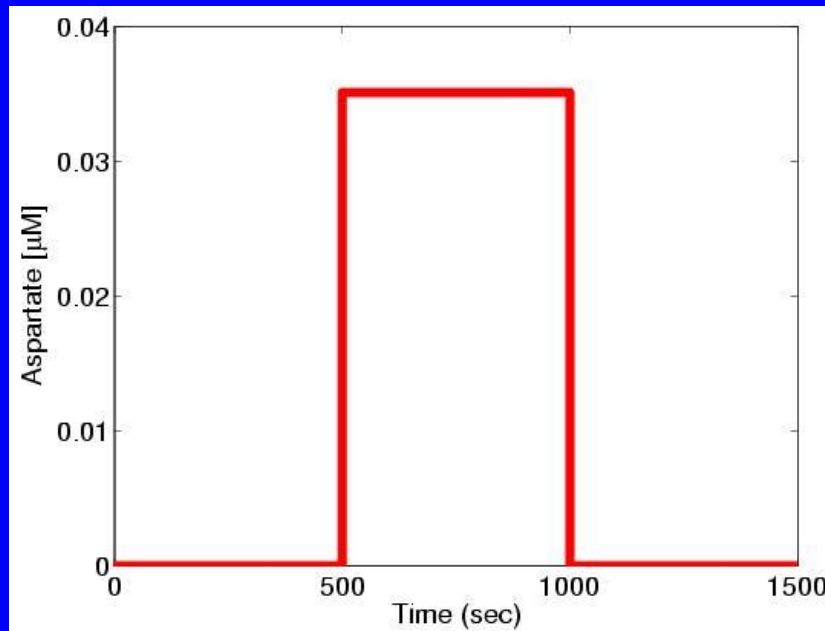
- Motor has a small range of sensitivity
- Cell is chemotactic for a large range of concentrations

⇒ System has to be adaptive:

Steady state of CheYp must be independent from absolute concentration of ligand

# Chemotaxis – The Task

**Input: Nutrient concentration    Output: Tumbling frequency**



**System performs a kind of differentiation**

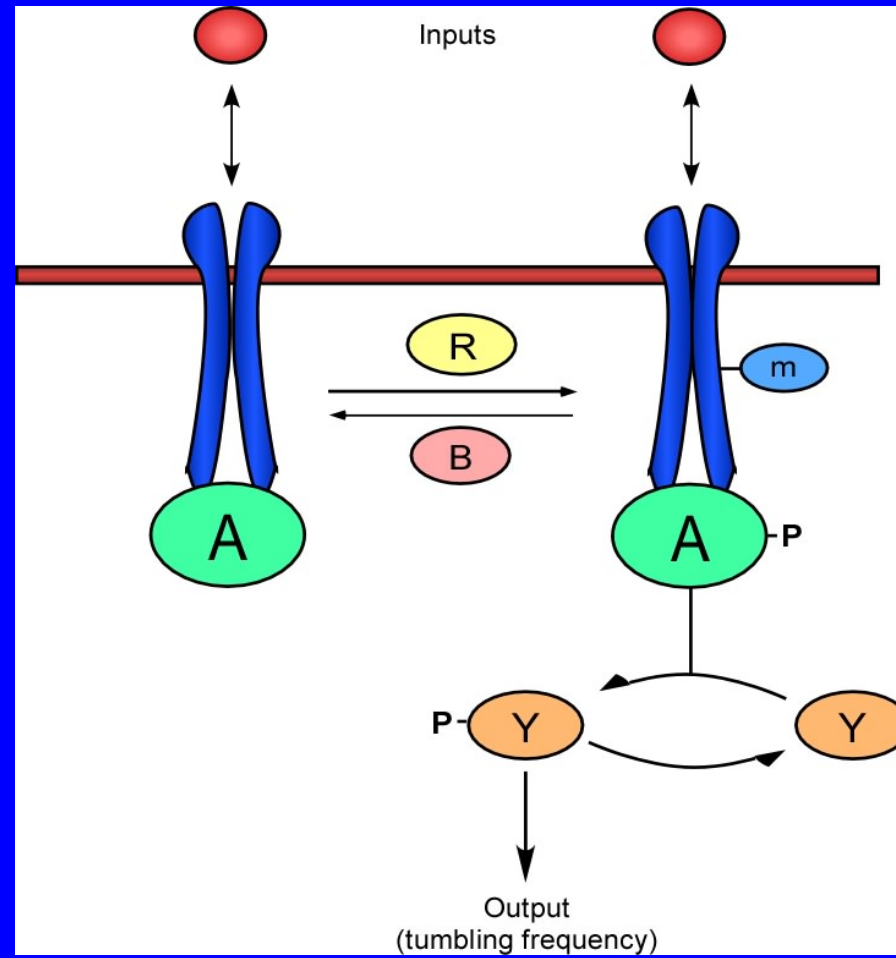
# The Players and their Roles

- **T: Receptors**
- **CheR: Methyltransferase, adds  $\text{CH}_3$**
- **CheB: Methyl-esterase, removes  $\text{CH}_3$**
- **CheA: Kinase, adds  $\text{PO}_4$**
- **CheZ: Phosphatase, removes  $\text{PO}_4$**
- **CheY: Signaling protein**

**Phosphorylation, Methylation = Chance of state**



# Barkai/Leibler Model – Graphical Version



# Barkai/Leibler Model – Mathematical Version

Probability for activating methylated receptor by ligand  $L$ :

$$p = \left(1 - \frac{L}{K_L + L}\right)$$

Concentration of activated receptors  $T_a$  :

$$T_a = p T_m$$

Methylation/demethylation dynamics of receptors:

$$\dot{T}_m = k_R R - k_B B \frac{T_a}{K_B + T_a}$$

Dynamics of  $A_p$ :

$$\dot{A}_p = k_A (A_{tot} - A_p) T_a - k_Y A_p (Y_{tot} - Y_p)$$

Dynamics of  $Y_p$ :

$$\dot{Y}_p = k_Y A_p (Y_{tot} - Y_p) - \gamma_Y Y_p$$

# Perfect Adaptation

Steady state of  $T_a$  from

$$\dot{T}_m = k_R R - k_B B \frac{T_a}{K_B + T_a} = 0$$

yields

$$T_a^{ss} = K_B \frac{k_R R}{k_B B - k_R R}$$

- Independent from ligand concentration  $L$
- Steady state is stable
- The same holds for  $Y_p$

Barkai & Leibler, Nature 387:913, 1997

## The Mechanism: $T_a = p(L) T_m(T_a)$

- Increasing  $L$  leads to fast decrease of  $T_a$
- $A_p$  &  $Y_p$  are fastly dephosphorylated
- $T_m$  is slowly increased
- Turns  $T_a$  and  $A_p$  &  $Y_p$  back to steady state
- Integral negative feedback control

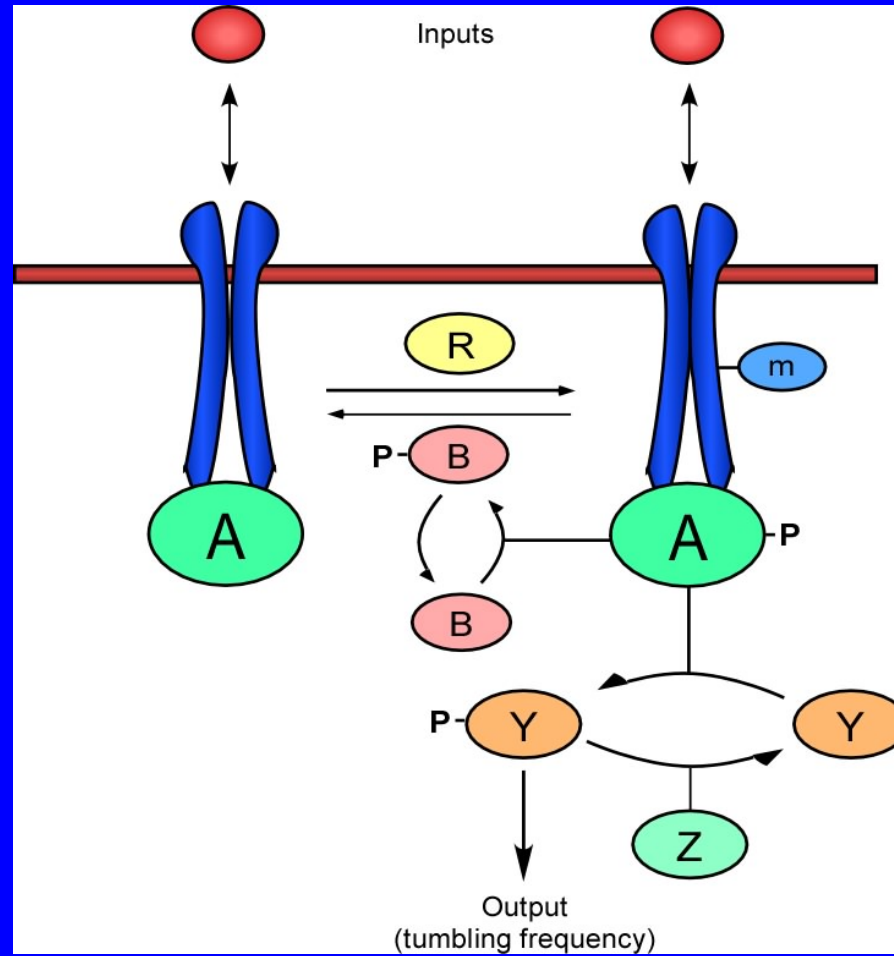
In words:

Degree of methylation compensates/remembers absolute concentration of ligand

**But ...**

**... this model is not realised by nature**

# Nature's E. Coli



# Sources of Variability

- **Intrinsic noise**

Differences between identical reporters within one cell

- Stochasticity of reactions

- **Extrinsic noise**

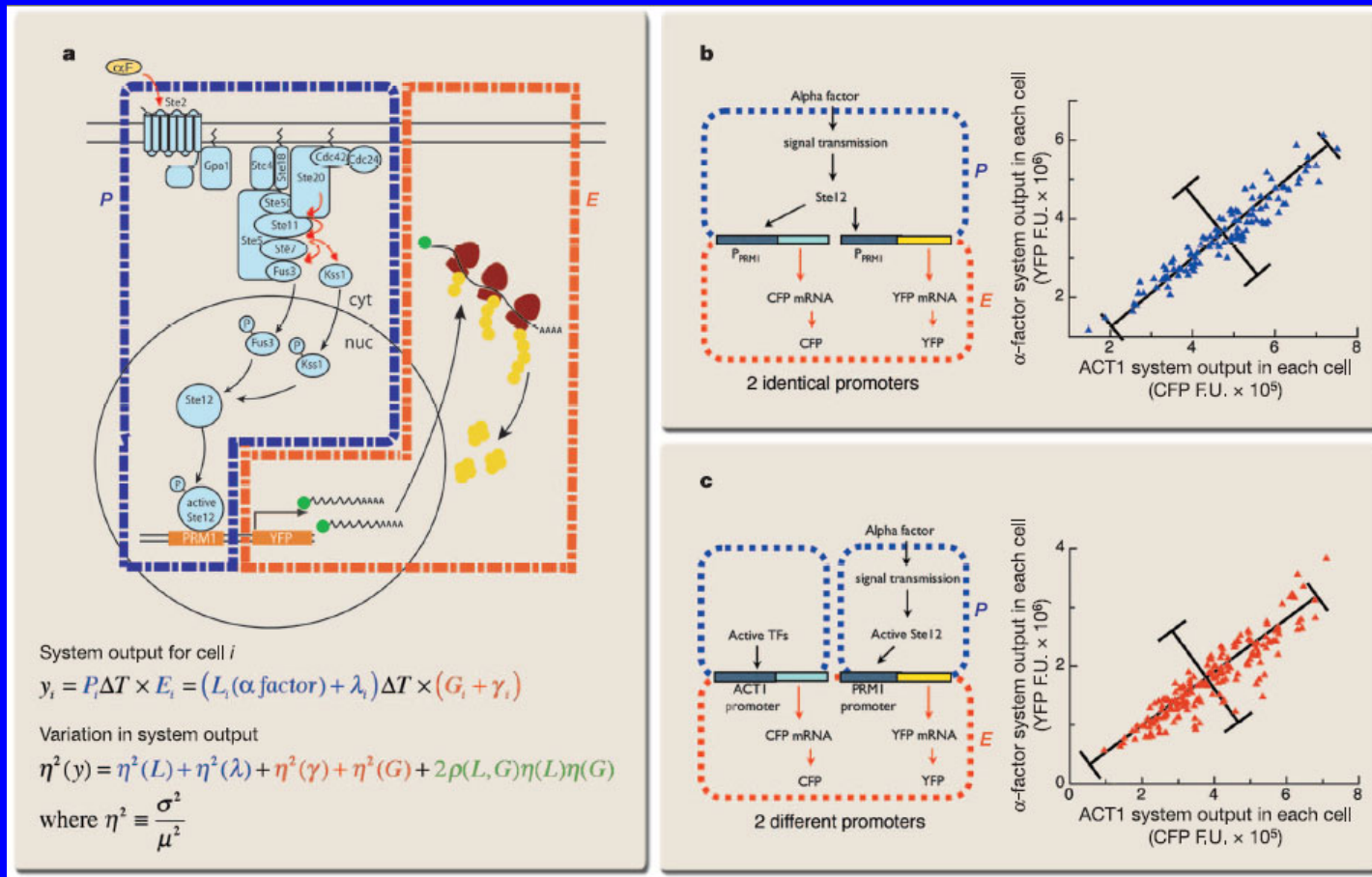
Differences between identical reporters in different cells

- Expression level of signaling proteins

- Number of ribosomes

Cell-to-cell variability

# Quantification of Variability



Colman-Lerner et al. Nature 437:699, 2005

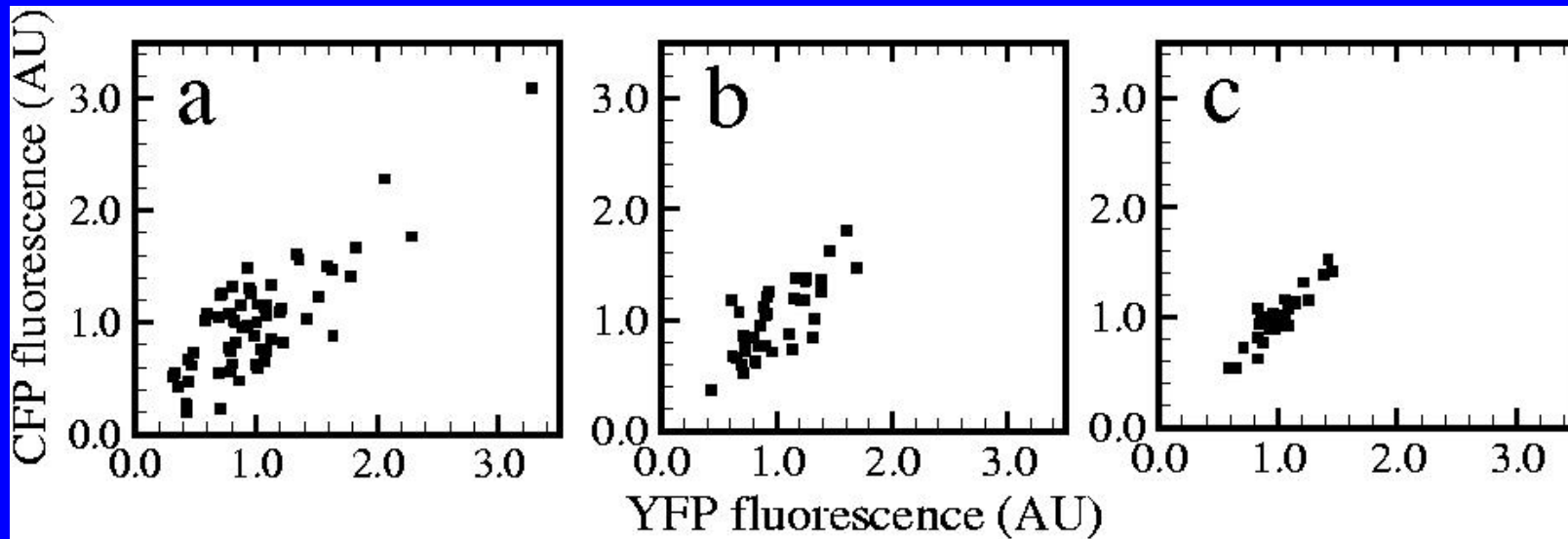


# Results

**E. coli and yeast:**

- **Extrinsic noise is larger than intrinsic noise**
- **Protein concentrations fluctuate in a correlated manner**

# Fluctuations and Chemotaxis



- Cell-to-cell fluctuations up to factor of ten
- Correlated fluctuations are dominant

# A Robustness Principle

The functionality of a pathway must be robust against fluctuations of protein levels.

For chemotaxis:

- Steady state level  $Y_p$  in  $[2.2 \mu\text{M}, 4.3 \mu\text{M}]$

- For correlated fluctuation:

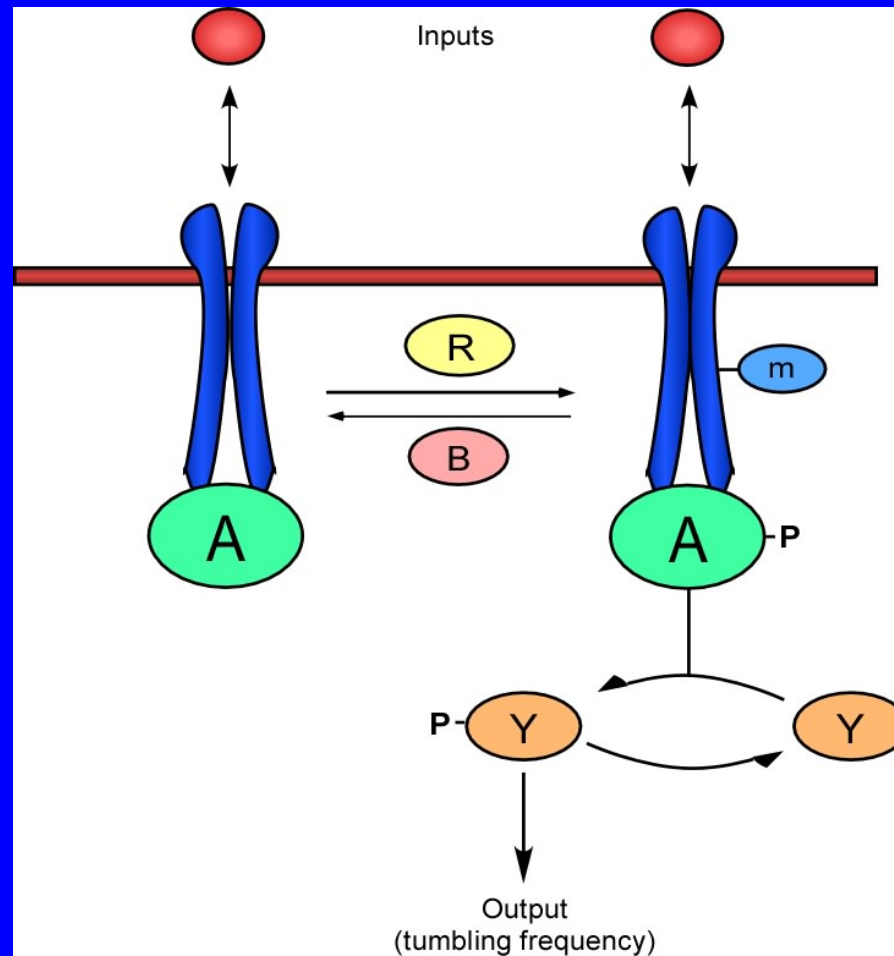
Steady state invariant under transformation:  $X_i \rightarrow \lambda X_i$

Important quantities may only depend on ratios of concentrations

- For uncorrelated fluctuations:

Use feedback-loops to attenuate noise

# Application to Barkai/Leibler Model



## Robustness of Barkai/Leibler Model

Steady states (with some approximations):

$$T_a^{ss} = K_B \frac{k_R R}{k_B B - k_R R} \quad \text{o.k.}$$

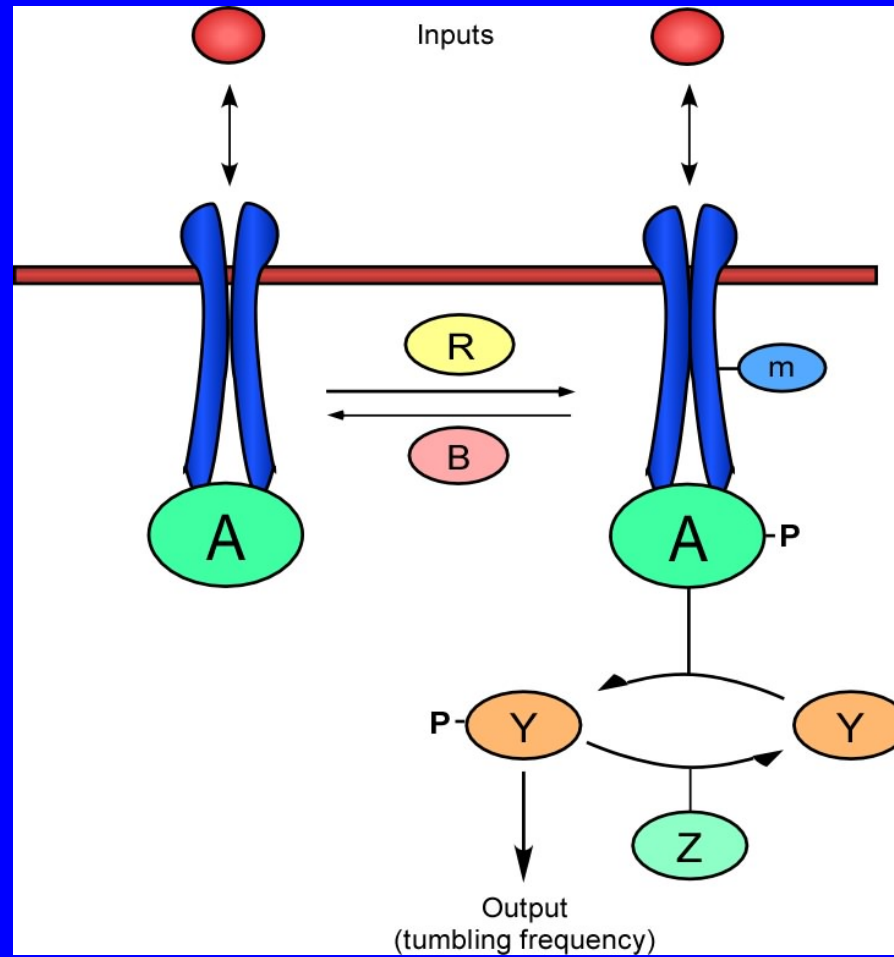
$$Ap^{ss} \approx \frac{k_A T_a^{ss}}{k_Y} \frac{A_{tot}}{Y_{tot}} \quad \text{o.k.}$$

$$Yp^{ss} = \frac{k_y Ap^{ss}}{k_Y Ap^{ss} + \gamma_Y} Y_{tot} \quad \text{not o.k.}$$

Cure:  $Yp$  must have a phosphatase (*CheZ*)

$$Yp^{ss} = \frac{k_y Ap^{ss}}{k_Z} \frac{Y_{tot}}{Z_{tot}} \quad \text{o.k.}$$

# Extension of the Model



## Robustness Against Correlated Fluctuations

- $Y_p$  must have a phosphatase ( $CheZ$ )
- Methyltransferase  $CheR$  has to work at saturation
- The pathway must be weakly activated,  $X_p \ll X_{tot}$

# Robustness Against Uncorrelated Fluctuations

Diminish uncorrelated noise by a classical feedback

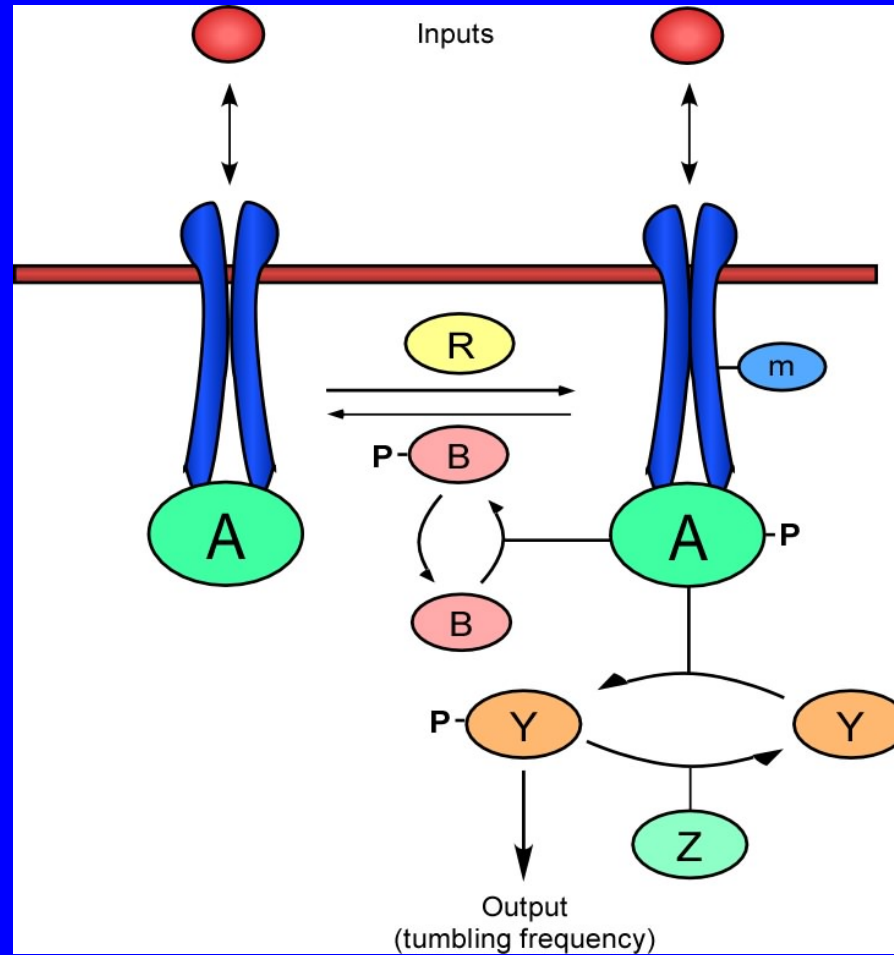
- Methyltransferase  $B$  can be phosphorylated by  $A_p$
- Only  $B_p$  can demethylate receptors

$$\Delta Y_p = -\frac{\frac{\partial f}{\partial T_a} \frac{\partial T_a}{\partial R}}{\alpha + \beta \frac{\partial B_p}{\partial A_p}} \Delta R$$

- Robustness against correlated fluctuations:  
 $\implies B_p$  must not have a phosphatase



# Final Model



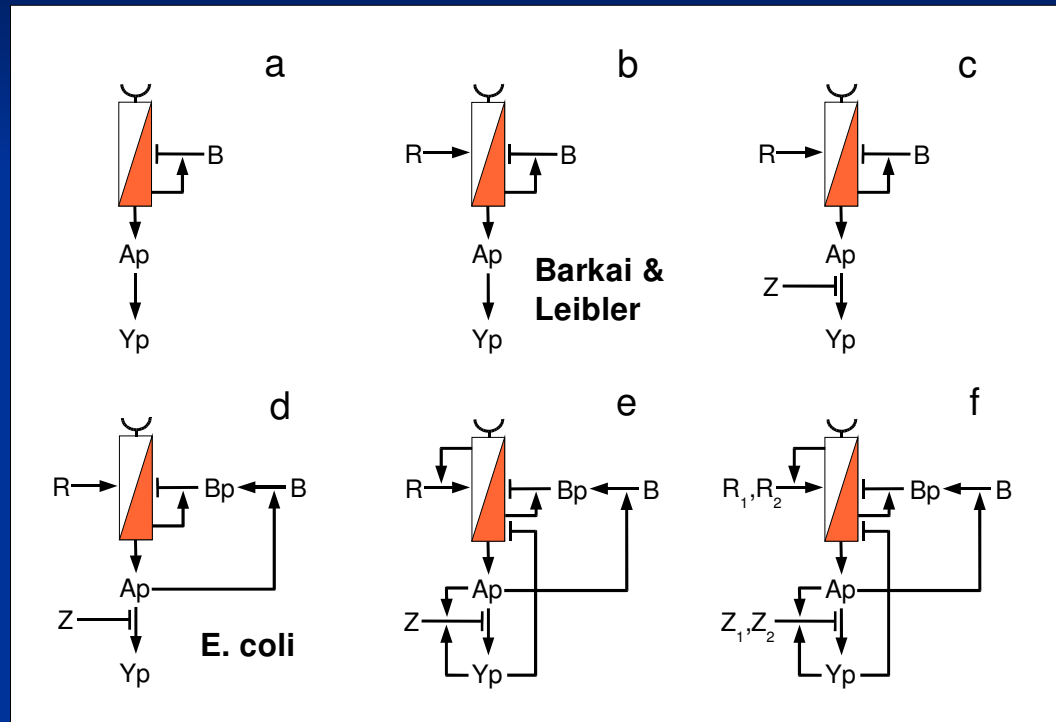
And this is how *E. coli* looks like

## *In silico* **Biology**

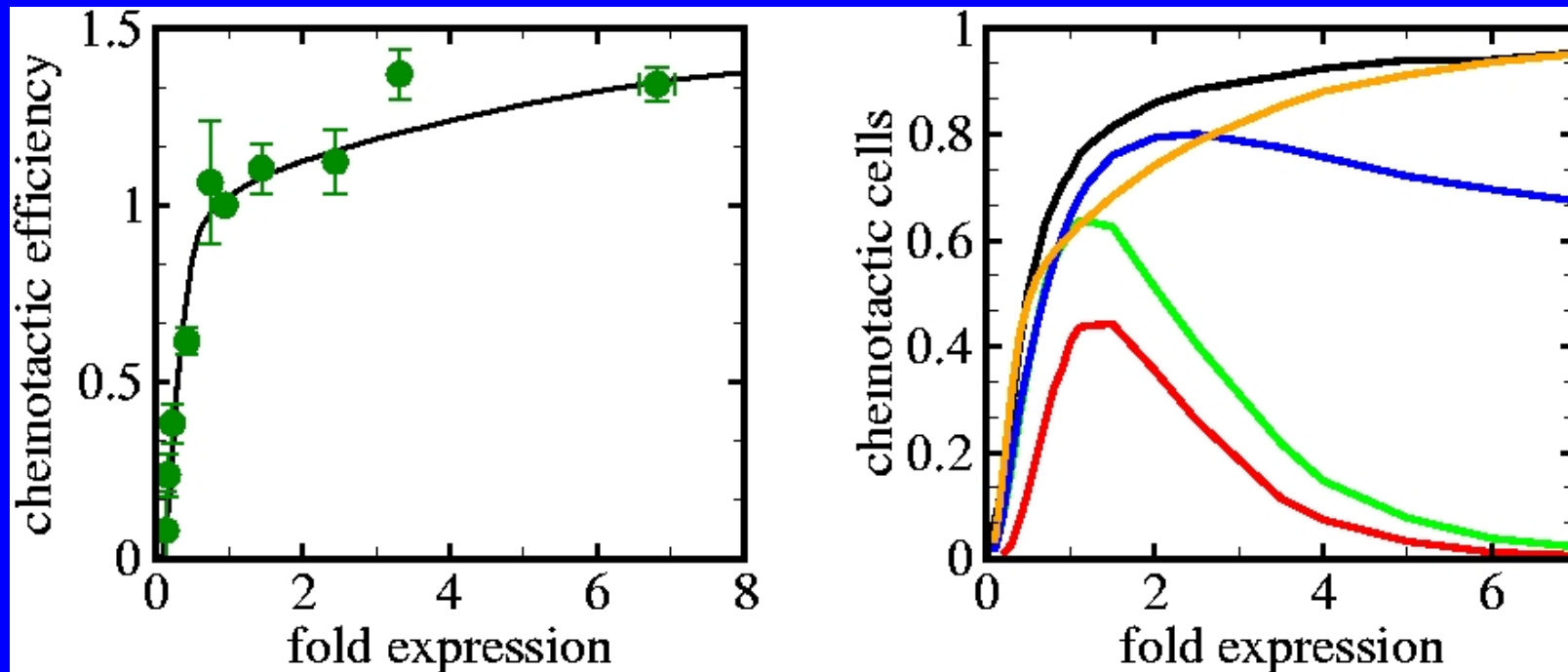
- **Choose different pathway topologies**
- **Parameters known experimentally**
- **Protein concentrations from experimental distributions**

**Compare chemotactic behaviour of *in silico* mutants to *E. coli* for different expression levels of proteins**

# Cartoons of Perfect Adaptive Pathways



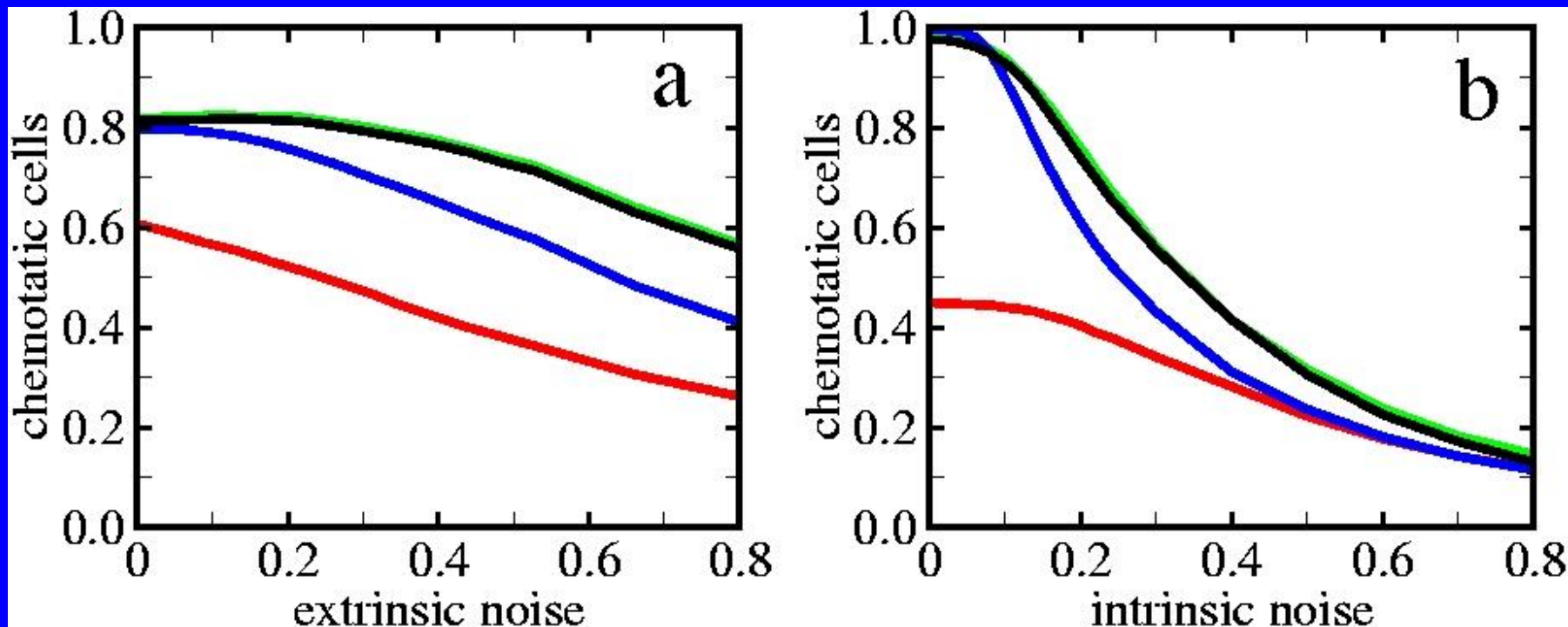
## Results: in vivo vs. in silico



red: Barkai/Leibler, black: final model, cyan: without feedback

blue: CheR not in saturation, green: CheBp with phosphatase

# Impossible Experiments



wild type: 0.4

wild type: 0.2

red: BL, black: fm, blue: w/out fb, green: mcm

## Conclusions

- **E. coli has to be adaptive and robust**
- **E. coli seems to be optimised to deal with fluctuations:**
  - **Uncorrelated noise: Feedback control**
  - **Correlated noise: Phosphatase here, saturation there**
- **E. coli is as complex as necessary but as simple as possible**

## Work done by

Physics Institute  
University of Freiburg

Centre for Molecular Biology  
University of Heidelberg

Markus Kollmann

Victor Sourjik

Kilian Bartholomé

Linda Lovdok

M. Kollmann, L. Lovdok, K. Bartholomé, J. Timmer, V. Sourjik.

Design principles of a bacterial signalling network, Nature 438:504, 2005

# Open Positions

- **BMBF Systems Biology of Hepatocytes** *HepatoSys*
- **DFG Graduate College 1305: Plant Signaling Systems**
- **BMBF Research Unit Systems Biology** *FRISYS*

[jeti@fdm.uni-freiburg.de](mailto:jeti@fdm.uni-freiburg.de)